# Cat # SL100468 Store at

# LipoJet™ In Vitro DNA and siRNA Transfection Kit (Ver. II)

----- A General Protocol for Transfecting
Mammalian Cell

| \$ | <b>SignaGen</b><br>Laboratories | ® |
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|    | Laboratories                    |   |

9601 Medical Center Drive Rockville MD 20850 FAX. 301-560-4919

Toll Free. 1-(866)-918-6812 Email: <u>info@signagen.com</u> Web: <u>www.signagen.com</u>

TEL. 301-330-5966



This product is for laboratory research ONLY and not for diagnostic use

### Introduction:

Based on our innovative and proprietary lipid-conjugation technology, LipoJet™ Transfection Kit, formulated from novel fluorinated cationic lipids, exhibits significant difference from other lipids transfection reagents in the market. LipoJet™ Transfection Kit is the most powerful yet very gentle gene delivery tool for a variety of applications including plasmid DNA and/or siRNA for most of mammalian cell types. Compared with leading products in the market, LipoJet™ is more cost-effective and always provides higher transfection efficiency with less cytotoxicity.

### Contents Per Kit:

- 1. 1 x 1.0 ml of LipoJet™ DNA In Vitro Transfection Reagent
- 2. 1 x 8.0 ml of LipoJet™ Transfection Buffer (5x)

### Important Guidelines for Transfection:

- LipoJet™ reagent was formulated for DNA and siRNA transfection. The following standard protocol is given for DNA and siRNA transfection to mammalian cells. For a protocol of siRNA/DNA cotransfection, please email us at <a href="mailto:info@signagen.com">info@signagen.com</a>
- For better efficiency, choosing LipoJet™ Transfection Buffer (1x ) is
- To lower cytotoxicity, transfect cells in presence of serum (10%) and antibiotics.

## Part I. A General Protocol for DNA Transfection.

# Step I. Preparation of Working Solution of LipoJet™ Transfection Buffer (1x )

 $\label{limited} \mbox{LipoJet}^{\mbox{\tiny M}}\mbox{ Transfection Buffer (5x ) is provided as 5x concentrated} \\ \mbox{stock solution.} \mbox{ To make working solution, dilute one part of the stock solution with 4 parts of $ddH_2O$. The LipoJet$^{\mbox{\tiny M}}$ Transfection Buffer (1x ) working solution is stable at RT for 24 months.$ 

Note: Always keep LipoJet  $^{\mathbb{M}}$  Transfection Buffer (5x ) at RT. If refrigerated, white precipitates may appear. It won't affect the transfection efficiency. After dilution with 4 parts of  $ddH_2O$  to make LipoJet  $^{\mathbb{M}}$  Transfection Buffer (1x ) working solution, the white precipitates will disappear.

## Step II. Cell Seeding:

Cells should be plated 18 to 24 hours prior to transfection so that the monolayer cell density reaches to the optimal 60-70% confluency at the time of transfection. Complete culture medium with serum and antibiotics is freshly added to each well 30-60 minutes before transfection.

**Note:** High serum levels (>5%) with antibiotics do NOT have inhibitory effect on transfection efficiency. We recommend using complete serum/antibiotics-containing medium to grow the cells during transfection.

# Step III. Preparation of LipoJet™ -DNA Complex and Transfection Procedures:

For different cell types, the optimal ratio of LipoJet<sup> $\mathbb{M}$ </sup> ( $\mu$ L):DNA ( $\mu$ g) varies from 2:1 to 3:1. We recommend using LipoJet<sup> $\mathbb{M}$ </sup> ( $\mu$ L):DNA ( $\mu$ g) at 2:1 at a starting point.

The following protocol is given for transfection in 24-well plates, refer to <u>Table 1</u> for transfection in other culture formats.

- For each well of 24-well plate, dilute 0.5 µg of DNA into 50 µl of LipoJet™ Transfection Buffer (1x ) prepared from <u>Step I</u>. Mix by vortexing.
- Add 1.0 µl of LipoJet™ reagent, vortex briefly to mix.
- Incubate for ~10 min at RT to allow LipoJet™/DNA complex to form.

**Note:** Never keep the LipoJet™/DNA complex longer than 20 min.

- Add the LipoJet™/DNA transfection mix to the cells in serum containing medium drop wise.
- Swirl plate gently to homogenize.
- Check transfection efficiency 24 to 48 hours post transfection. 48 hours usually give better efficiency.

Table 1. Recommended Amounts for Different Culture Vessel Formats

| Culture Dish | Culture<br>Medium<br>(ml) | Plasmid<br>DNA<br>(µg) | LipoJet™<br>Transfection<br>Buffer (1x )<br>(µL) | LipoJet™<br>Reagent<br>(μL) |
|--------------|---------------------------|------------------------|--|-----------------------------|
| 96-well      | 0.1                       | 0.1                    | 5  | 0.2 ~ 0.3                   |
| 70-WEIL      | 0.1                       | 0.1                    |  | 0.2 ~ 0.3                   |
| 48-well      | 0.25                      | 0.25                   | 25   | 0.5 ~ 0.75                  |
| 24-well      | 0.5                       | 0.5                    | 50   | 1 ~ 1.5                     |
| 6-well       | 2                         | 2.0                    | 200  | 4 ~ 6                       |
| 35 mm dish   | 2                         | 2.0                    | 200  | 4 ~ 6                       |
| 60 mm dish   | 4                         | 4.0                    | 400  | 8 ~ 12                      |
| 10 cm / T75  | 10                        | 10                     | 800  | 20 ~ 30                     |
| 15 cm / T175 | 20                        | 20                     | 1600   | 40 ~ 60                     |

Storage: LipoJet  $^{\mathtt{m}}$  Reagent is stable for up to 12 months at +4  $^{0}\text{C}$  after receipt

# Cat # SL100468 Store at 4 <sup>o</sup>

# LipoJet™ In Vitro DNA and siRNA Transfection Kit

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| €             | SignaGen®<br>Laboratories |
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### Part II. A General Protocol for siRNA Transfection.

# Step I. Preparation of Working Solution of LipoJet™ Transfection Buffer

LipoJet<sup>™</sup> Transfection Buffer (5x ) is provided as 5x concentrated stock solution. To make working solution, dilute one part of the stock solution with 4 parts of  $ddH_2O$ . The 1x LipoJet<sup>™</sup> Transfection Buffer is stable at RT for 24 months.

Note: Always keep LipoJet  $^{\mathbb{M}}$  Transfection Buffer (5x ) at RT. If refrigerated, white precipitates may appear. It won't affect the transfection efficiency. After dilution with 4 parts of  $ddH_2O$  to make LipoJet  $^{\mathbb{M}}$  Transfection Buffer (1x ) working solution, the white precipitates will disappear.

### Step II. Cell Seeding:

Cells should be plated 18 to 24 hours prior to transfection so that the monolayer cell density reaches to the optimal 50% confluency at time of transfection. Complete culture medium with serum and antibiotics is freshly added to each well 30-60 minutes before transfection.

Note: High serum levels (>5%) with antibiotics usually do not have inhibitory effect on transfection efficiency. We recommend using complete serum/antibiotics-containing medium to grow the cells during transfection.

# Step III. Preparation of LipoJet™-siRNA Complex and Transfection Procedures:

For optimal siRNA-mediated silencing, we recommend using 10-80 nM siRNA (final concentration). The following protocol is given for transfection in 6-well plate, refer to <u>Table 2</u> for transfection in other culture formats.

- For each well, dilute 20 ~ 160 pmoles siRNA (for a final concentration of 10 to 80 nM per well) into 200 μl of LipoJet™ Transfection Buffer (1x) prepared from <u>Step I</u>. Mix gently.
- Add 4 µl of LipoJet™ reagent, vortex briefly to mix.
- Incubate for ~10 min at RT to allow LipoJet™/siRNA complexes to form

Note: Never keep the LipoJet™/siRNA complex longer than 20 min.

- Add the LipoJet™/siRNA transfection mix to the cells in serumcontaining medium drop wise.
- Swirl plate gently to homogenize.
- Check siRNA silencing efficiency 24 to 72 hours post transfection. 48-72 hours usually give better efficiency.

# Table 2. Recommended Amounts for Different Culture Vessel

| Tornics         |                           |                               |   |                             |  |
|-----------------|---------------------------|-------------------------------|---|-----------------------------|--|
| Culture<br>Dish | Culture<br>Medium<br>(ml) | siRNA<br>(pmoles)<br>10~80 nM | LipoJet™<br>Transfection Buffer<br>(1x ) (μL) | LipoJet™<br>Reagent<br>(μL) |  |
| 96-well         | 0.1                       | 1 ~ 8                         | 10  | 0.4                         |  |
| 48-well         | 0.25                      | 2.5 ~ 20                      | 25  | 1                           |  |
| 24-well         | 0.5                       | 5 ~ 40                        | 50  | 2                           |  |
| 6-well          | 2                         | 20 ~ 160                      | 200   | 4                           |  |
| 35 mm dish      | 2                         | 20 ~ 160                      | 200   | 4                           |  |
| 60 mm dish      | 4                         | 40 ~ 320                      | 400   | 8                           |  |
| 10 cm /<br>T75  | 10                        | 100 ~ 800                     | 800   | 20                          |  |